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# $\beta$ -Tricalcium phosphate nanofiber scaffolds with fine unidirectional grains



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## ABSTRACT

$\beta$ -Tricalcium phosphate (TCP) is an osteoinductive and resorbable material that offers superior biochemical activity for bone regeneration therapies. TCP nanofiber scaffolds are particularly promising owing to their high porosity and chemical composition that resemble the mineral phase of natural extracellular matrix. TCP nanofiber scaffolds were fabricated by electrospinning. The nanofiber morphology promoted unidirectional grain growth during the sintering stage due to lack of surrounding grains in the other two principal directions. The average fiber diameter after sintering was 100–125 nm, which is the smallest grain size ever reported for TCP.

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## 1. Introduction

Nanofiber materials show enhanced structural and functional properties compared to bulk materials, and have been applied in catalysis, energy, biomedicine, sensing and electronics [1–4]. In the field of biomedicine, nanofiber materials are widely used as scaffolds for tissue regeneration. Nanofibers are often made by electrospinning, which involves applying a high voltage between a hollow needle that releases a solution and a counter electrode. The electrified solution is dragged into a fiber shape that dries during transport and is collected on the counter electrode.

Bioceramic materials have been extensively used to repair bone defects [5–7]. They offer similar chemical and mechanical properties as bone [8–10]. Bioactive ceramics, e.g. calcium phosphates and bioactive glasses, have been successfully applied [5,6,11,12]. The  $\beta$ -phase of tricalcium phosphate ( $\beta$ -TCP) has shown excellent performance as a scaffold for bone regeneration [11–14]. It offers osteogenic induction and is resorbable by the body [11,14,15].

$\beta$ -TCP is a much less reported phase than hydroxyapatite (HA) or calcium deficient HA (CaDHA).  $\beta$ -TCP results from decomposition of CaDHA at high temperature [14,16]. CaDHA forms a secondary phase of  $\beta$ -TCP above 700 °C.  $\beta$ -TCP is normally sintered above 1050 °C, but such high temperatures induce grain growth [5].

Porous  $\beta$ -TCP with small grain sizes has superior properties in terms of osteogenic differentiation, inducing bone growth even in soft tissues [12,14,17]. Bioglass [3,18,19] and HA [20,21] nanofibers by electrospinning have been reported. However, to the best of our knowledge no attempt to fabricate  $\beta$ -TCP nanofibers has been reported. Here, we demonstrate the successful fabrication of  $\beta$ -TCP nanofibers by electrospinning.

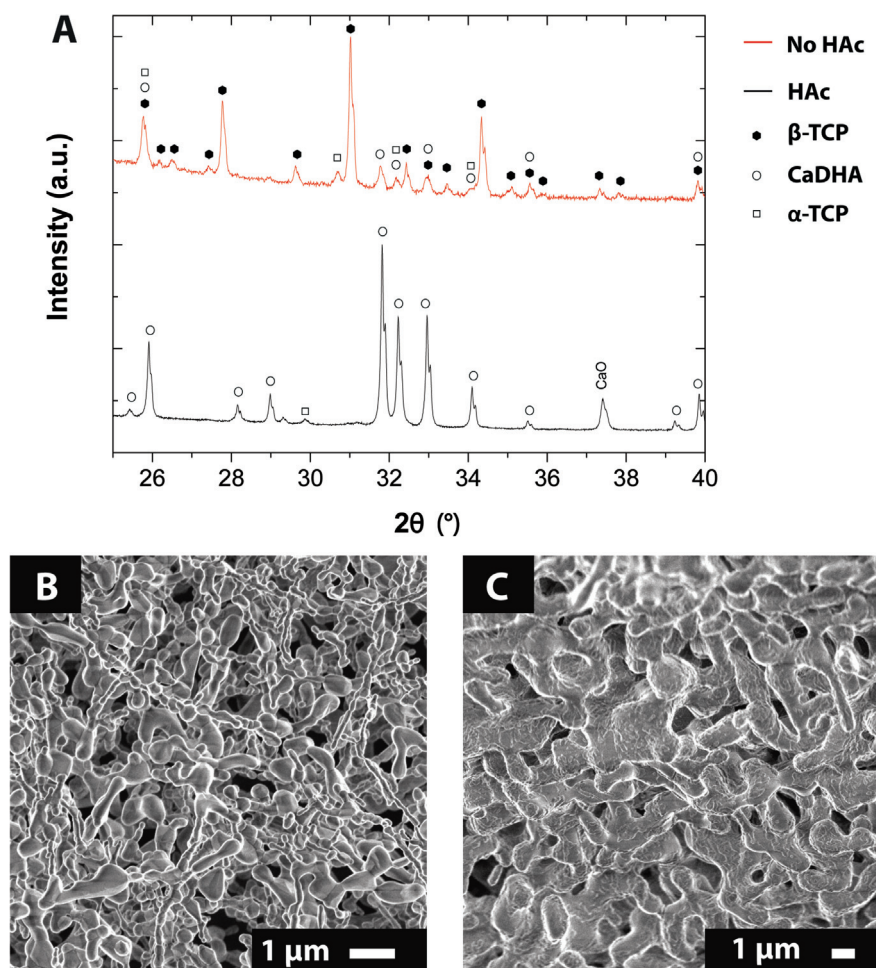
## 2. Materials and methods

TCP was synthesized using an acid-base reaction between calcium nitrate tetrahydrate (Sigma Aldrich) and phosphoric acid (Sigma Aldrich). A 1 M calcium nitrate tetrahydrate solution was prepared in 1:1 vol/vol ethanol:water. A separate 0.67 M phosphoric acid solution was prepared in 4:1 vol/vol ethanol:water. The solutions were stabilized by adding a third solution consisting of either (i) acetic acid (HAc, Sigma Aldrich) acting as chelating agent, or (ii) 1:1 vol/vol ethanol:water to decrease the ion concentration. The three solutions were mixed in a volume ratio 2:2:1. Polyvinylpyrrolidone (PVP k90; Mw 360,000 g/mol) was added at a concentration of 100 mg/ml to control the viscosity.

Electrospinning was performed at 25 °C and relative humidity 30%. The inner diameter of the spinneret was 0.8 mm. The distance between spinneret and Al collector plate was 17 cm. The flow rate was 0.25–0.6 mL/h and the voltage 12–15 kV. The samples were annealed in a convection oven (Eurotherm) or in a microwave oven (Milestone MicroSynth) at 950–1300 °C. The dwell time was

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**Fig. 1.** (A) XRD patterns of samples spun with and without acetic acid as stabilizer. The samples were annealed at 1050 °C. (B) SEM image of a sample spun without using acetic acid and annealed at 1050 °C. (C) SEM image of a sample spun without acetic acid and annealed at 1300 °C. The scale bars are 1  $\mu\text{m}$ .

0–1.5 h, using heating and cooling rates of 5–20 °C/min. A Zeiss Merlin Scanning electron microscope (SEM) was used to investigate the microstructure of the prepared material. A PANalytical X'Pert X-ray Diffraction (XRD) machine was used for phase analysis and grain size determination using the Scherer equation. The XRD data were analyzed with High Score Plus<sup>®</sup>. Fiber diameters and grain sizes were determined by statistical analysis of SEM images.

### 3. Results and discussion

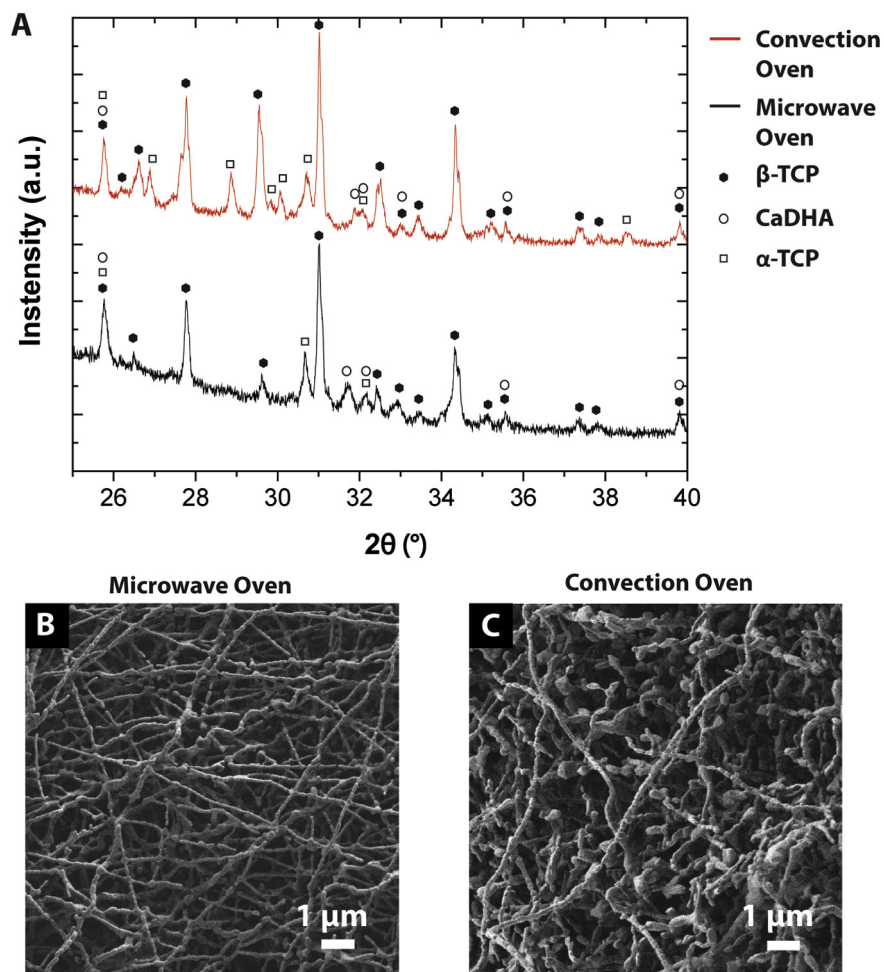
Solutions for electrospinning are normally weak electrolytes with low conductivity [22]. Conductive metal salt solutions as used here require adaptation to limit the effective ionic strength, and facilitate the stability of the electrospinning process. We attempted HAc as solvent and chelating agent for the metal ions, but a CaDHA phase was obtained instead of  $\beta$ -TCP (Fig. 1A, bottom). When we used ethanol/water to just dilute the solutions,  $\beta$ -TCP was formed (Fig. 1A, top). To keep the jet containing non-stabilized ionic charges stable [22], low voltage and flow rate were used.

Sintering of bulk  $\beta$ -TCP starts above 750 °C, and the highest densities are achieved above 1100 °C [5]. TCP is usually sintered at temperatures >1050 °C. However, when we annealed our samples at 1050–1300 °C, the resulting fibers were not continuous due to sintering effects (Fig. 1B,C). The microstructure resembled that of porous CaDHA [5,23]. High surface area nanofibers could be formed only at temperatures lower than 1000 °C.

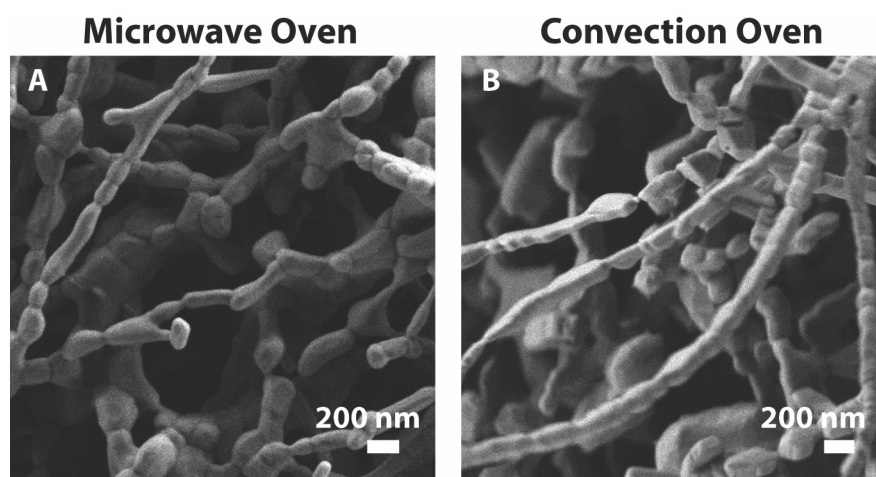
To induce crystallization while avoiding surface area reduction and/or grain growth, short thermal treatments are needed. We thus heated the nanofibers to 950 °C, and then cooled them down immediately with heating/cooling rates of 5–20 °C/min. In this case only CaDHA fibers were obtained.

When the dwell time was increased to 1.5 h,  $\beta$ -TCP was obtained as the main phase at 950 °C (Fig. 2A). However, HA and  $\alpha$ -TCP secondary phases were also formed, especially in the sample that had been annealed in a convection oven. Similar secondary phases have also been reported in previous studies on TCP formation [14,16]. The superior phase purity of the microwave-treated material is attributed to the effect of microwave radiation heating, which may allow better crystallization [24–27].

The samples that were thermally annealed in the microwave oven (Fig. 2B) kept its nanofiber morphology better than the ones annealed in a convection oven (Fig. 2C), which is attributed to the faster heating/cooling rates in the former experiment. The fiber diameters were  $120 \pm 20$  and  $120 \pm 30$  nm, respectively. The grains were approximately 100 nm wide (Fig. 3), which is close to the fiber diameter. This value is much smaller than normally reported micrometer grain sizes for  $\beta$ -TCP, and about 2.5 smaller than the smallest reported  $\beta$ -TCP grain sizes known to us [5]. The grains are anisotropic, directionally oriented along the long axis of the fibers. This is probably caused by the lack of mass transport towards the grain from the surrounding matrix during sintering.



**Fig. 2.** (A) XRD patterns of the samples shown in (B) and (C), respectively. (B) SEM image of a sample annealed in a microwave oven at 950 °C for 1.5 h and a heating/cooling rate of 20 °C/min. The scale bar is 1 μm. (C) SEM image of a sample annealed in a convection oven at 950 °C for 1.5 h and a heating/cooling rate of 10 °C/min. The scale bar is 1 μm.



**Fig. 3.** Formation of elongated grains. (A) Sample annealed in a microwave oven for 1.5 h at a heating/cooling rate of 20 °C/min. The scale bar is 200 nm. (B) Sample annealed in convection oven for 1.5 h at a heating/cooling rate of 10 °C/min. The scale bar is 200 nm.

#### 4. Conclusions

β-TCP nanofiber mats with only a minor fraction of secondary phase were successfully fabricated. The grain sizes were about

100 nm. Rapid heating/cooling and short sintering times help to maintain small grain sizes. The nanofiber morphology promoted unidirectional grain growth. β-TCP nanofiber mats may offer an excellent platform for bone regeneration studies.



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